Anti-ulcerogenic activity of *Gynura procumbens* leaf extract against experimentally-induced gastric lesions in rats

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Several plants are used in folk medicine to treat gastrointestinal disorders. *Gynura procumbens* is a medicinal plant commonly used in traditional treatment of many ailments. In this study, *G. procumbens* ethanolic leaf extract (GPELE) was used to investigate its gastroprotective effect in adult Sprague dawley rats which were divided into six groups. The rats were orally pre-treated with carboxymethyl cellulose (CMC) solution (ulcer control groups), omeprazole 20 mg/kg (reference group), 50, 100, 200 and 400 mg/kg of GPELE in CMC solution (experimental groups), one hour before oral administration of absolute ethanol to generate gastric mucosal injury. After an additional hour, the rats were sacrificed and the ulcer areas of the gastric walls were determined. The ulcer control group exhibited severe mucosal injury, whereas groups pre-treated with GPELE exhibited significant protection of gastric mucosal injury. These findings were also confirmed by histological studies. Acute toxicity study with a higher dose of 5 g/kg did not manifest any toxicological signs in rats. These results suggest that GPELE promotes ulcer protection as ascertained grossly by significant reduction of ulcer area, and histologically by comparatively decreases in ulcer areas, reduction or absence of edema and leucocytes infiltration of submucosal layer compared to ulcer control group.

**Key words:** *Gynura procumbens*, cytoprotection, gastric ulcer, histology, ethanolic extract.

**INTRODUCTION**

Gastric ulcer is an illness that affects a considerable number of people worldwide. The etiological factors of this disorder include: stress, smoking, nutritional deficiencies, infections, frequent and indiscriminate use of non-steroidal anti-inflammatory drugs (NSAIDs) (Khazaei and Salehi, 2006). The pathogenesis of gastro-duodenal ulcers is influenced by various aggressive and defensive factors, such as mucus secretion, mucosal barrier, acid-pepsin secretion, blood flow, cellular regeneration and endogenous protective agents (Mizui et al., 1987). Although the introduction of proton-pump inhibitors to the classic anti-ulcer therapy had revolutionized treatment of peptic ulcers and other gastrointestinal disorders, there is still no complete cure for this disease. It has been shown that long term use of these drugs may be associated with ineffectiveness of different drug regimens and even resistance to drugs are emerging (Al-Mofleh et al., 2007). Thus, there is an urgent need to identify more effective and safe anti-ulcer agents.

A widespread search has been launched to identify new anti-ulcer therapies from natural sources. Herbs, medicinal plants, spices, vegetables and crude drug substances are considered to be a potential source to combat various diseases including gastric ulcer. In the scientific literature, a large number of medicinal plants with gastric anti-ulcer potential have been reported (Al-Mofleh et al., 2008; Devi et al., 2008; Coelho et al., 2009). *G. procumbens* (Merr.) which is known in Malaysia as “Sambung nyawa.” is widely distributed in South East...

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Asian countries such as Indonesia, Malaysia, and Thailand. The plant is used in the traditional treatment of many ailments such as inflammation, rheumatism, viral diseases of skin, kidney diseases, rashes, fever, migraine, constipation, and cancer (Perry, 1980). The leaves of this plant are not toxic (Rosidah et al., 2009) and have been shown to possess anti-herpes simplex virus (Nawawi et al., 1999), anti-hyperglycemic (Akowuah et al., 2002), anti-inflammatory (Iskander et al., 2002), anti hyperlipidemic (Zhang and Tan, 2000) and anti-hypertensive effects (Kim et al., 2006).

Several studies have shown that GPELE contains several pharmaceutically active chemical constituents, such as flavonoids, saponins, tannins, and terpenoids (Akowuah et al., 2002). Thus far, there is no data available on gastroprotective activity of GPELE. The present study was undertaken to evaluate anti-ulcerogenic properties of GPELE in rats.

MATERIALS AND METHODS

In this study, omeprazole was used as the reference anti-ulcer drug, and was obtained from the University Malaya Medical Centre (UMMC) Pharmacy. The drug was dissolved in carboxymethyl cellulose (CMC) and administered orally to the rats in concentrations of 20 mg/kg body weight (5 ml/kg) according to the recommendation of Pedernera et al. (2006).

Plant specimen and extract preparation

G. procumbens leaves were obtained from Ethno Resources Sdn Bhd, Selangor Malaysia, and identified by comparison with the Voucher specimen deposited at the Herbarium of Rimba Ilmu, Institute of Science Biology, University of Malaya, Kuala Lumpur. The dried leaves were powdered using electrical blender. Hundred grams of the fine powder were soaked in 500 ml of 95% ethanol in conical flask for 3 days. After 3 days the mixture was filtered using a fine muslin cloth followed by filter paper (Whatman No. 1) and distilled under reduced pressure in an Eyela rotary evaporator (Sigma-Aldrich, USA).

The dry extract was then dissolved in CMC (0.25% w/v) and administered orally to rats in concentrations of 50, 100, 200 and 400 mg/kg body weight (5 ml/kg body weight) according to the recommendation of Pasquale et al. (1995).

Acute toxicity test LD₅₀

Adult male and female Sprague Dawley rats (6 - 8 weeks old) were obtained from the Animal House, Faculty of Medicine, University of Malaya, Kuala Lumpur (Ethics No. PM 07/05/2008 MAA (a) (R)). The rats weighed between 150 - 180 g. The animals were given standard rat pellets and tap water ad libitum. The acute toxicity study was used to determine a safe dose for the rhizome extract. Thirty six rats (18 males and 18 females) were assigned equally into 3 groups labeled as vehicle (CMC, 0.25% w/v, 5 ml/kg); 2 and 5 g/kg of GPELE preparation, respectively.

The animals fasted over-night (food but not water) prior to dosing. Food was withheld for a further 3 - 4 h after dosing. The animals were observed for 30 min and 2, 4, 8, 24 and 48 h after the administration for the onset of clinical or toxicological symptoms. Mortality, if any was observed over a period of 2 weeks. The acute toxicity LD₅₀ was calculated as the geometric mean of the dose that resulted in 100% lethality and that which caused no lethality at all. The animals were sacrificed on the 15th day. Histological, hematological and serum biochemical parameters were determined following standard methods (Bergmeyer, 1980; Tietz et al., 1983). The study was approved by the ethics committee for animal experimentation, Faculty of Medi-cine, University of Malaya, Malaysia. All animals received human care according to the criteria outlined in the “Guide for the Care and Use of laboratory Animals” prepared by the National Academy of Sciences and published by the national Institute of health.

Experimental animals

Sprague Dawley healthy adult male rats were obtained from the Experimental Animal House, Faculty of Medicine, University of Malaya, and Ethic No. PM/27/07/2009/MAA (R). The rats were divided randomly into 6 groups of 6 rats each. Each rat that weighed between 200 - 225 g was placed individually in a separate cage (one rat per cage) with wide-mesh wire bottoms to prevent coprophagia during the experiment. The animals were maintained on standard pellet diet and tap water. The study was approved by the Ethics Committee for Animal Experimentation, Faculty of Medicine, University of Malaya, Malaysia. Throughout the experiments, all animals received human care according to the criteria outlined in the “Guide for the Care and Use of laboratory Animals” prepared by the National Academy of Sciences and published by the national Institute of health.

Gastric ulcer-induction by ethanol

The rats fasted for 48 h before the experiment (Garg et al., 1993), but were allowed free access to drinking water up till 2 h before the experiment. Gastric ulcer was induced by orogastric intubation of absolute ethanol (5 ml/kg) according to the method described by De Pasquale et al. (1995) with slight modification. Ulcer control groups were orally administered vehicle (CMC, 0.25% w/v, 5 ml/kg). The reference group received oral doses of 20 mg/kg omeprazole in CMC (5 ml/kg) as positive control.

Experimental groups were orally administered GPELE in CMC solution (5 ml/kg) at doses of 50, 100, 200 and 400 mg/kg. One hour after this pre-treatment all groups of rats were lavaged with absolute ethanol (5 ml/kg) in order to induce gastric ulcers. The rats were euthanized by cervical dislocation 60 minutes later (Paiva et al., 1998) under an overdose of diethyl ether anesthesia and their stomachs were immediately excised.

Gross gastric lesions evaluation

Ulcers of the gastric mucosa appear as elongated bands of hemorrhagic lesions parallel to the long axis of the stomach. Gastric mucosa of each rat was thus examined for damage. The length and width of the ulcer (mm) were measured by a planimeter (10 x 10 mm² = ulcer area) under dissecting microscope (1.8x). The ulcerated area was measured by counting the number of small squares, 2 x 2 mm, covering the length and width of each ulcer band.

The sum of the areas of all lesions for each stomach was applied to the calculation of the ulcer area (UA) wherein the sum of small squares x 4 x 1.8 = UA (mm²) as described by Kauffman and Grossman (1978) with slight modification. The inhibition percentage (I.O %) was calculated by the following formula as described by Njar et al. (1995) with slight modification.

\[ I.O \% = \left( \frac{UA_{\text{control}} - UA_{\text{treated}}}{UA_{\text{control}}} \right) \times 100\% . \]
Table 1. Effect of GPELE on ulcer area and inhibition percentage in rats.

<table>
<thead>
<tr>
<th>Animal group</th>
<th>Pre-treatment (5 ml/kg dose)</th>
<th>Ulcer area (mm$^2$) (Mean ± S.E.M)</th>
<th>Inhibition (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>CMC (Ulcer control)</td>
<td>984.17 ± 5.39$^a$</td>
<td>-</td>
</tr>
<tr>
<td>2</td>
<td>Omeprazole (20 mg/kg)</td>
<td>180.00 ± 9.66$^b$</td>
<td>81.71</td>
</tr>
<tr>
<td>3</td>
<td>GPELE (50 mg/kg)</td>
<td>391.67 ± 13.02$^c$</td>
<td>60.20</td>
</tr>
<tr>
<td>4</td>
<td>GPELE (100 mg/kg)</td>
<td>225.00 ± 5.92$^d$</td>
<td>77.14</td>
</tr>
<tr>
<td>5</td>
<td>GPELE (200 mg/kg)</td>
<td>115.00 ± 7.64$^e$</td>
<td>88.35</td>
</tr>
<tr>
<td>6</td>
<td>GPELE (400 mg/kg)</td>
<td>55.83 ± 5.39$^f$</td>
<td>94.33</td>
</tr>
</tbody>
</table>

All values are expressed as mean ± standard error mean. Means with different superscripts are significantly different. The mean difference is significant at the 0.05 level.

Figure 1a. Gross appearance of the gastric mucosa in a rat pre-treated with 5 ml/kg of CMC (negative control). Severe injuries are seen in the gastric mucosa.

Figure 1b. Gross appearance of the gastric mucosa in a rat pre-treated with 5 ml/kg of omeprazole (20 mg/kg). Injuries to the gastric mucosa are milder compared to the injuries seen in the negative control rat.

Histological evaluation of gastric lesions

Specimens of the gastric walls from each rat were fixed in 10% buffered formalin and processed in a paraffin tissue processing machine. Sections of the stomach were made at a thickness of 5 µ and stained with hematoxylin and eosin for histological evaluation.

Statistical analysis

All values were reported as mean ± S.E.M. The statistical significance of differences between groups was assessed using one-way ANOVA. A value of p < 0.05 was considered significant.

RESULTS

Acute toxicity study

Animals treated with GPELE at a dose of 2 and 5 g/kg were kept under observation for 14 days. All the animals remained alive and did not manifest any significant sign of toxicity at these doses. There were no abnormal signs, behavioral changes, body weight changes, or macroscopic finding at any time of observation. There was no mortality in the above-mentioned doses at the end of 14 days of observation. Histological examination of liver and kidney, hematology and serum biochemistry revealed no significant differences between the different groups. From these results it is concluded that the extract is quite safe even at these higher doses and has no acute toxicity and the oral lethal dose (LD$_{50}$) for the male and female rats were greater than 5 g/kg body weight.

Gross evaluation of gastric lesions

The anti-ulcer activity of GPELE in ethanol-induced gastric lesion model is shown in Table 1. Results showed that rats pre-treated with GPELE extracts before being given absolute alcohol had significantly reduced areas of gastric ulcer formation compared to rats pre-treated with only CMC (ulcer control group) (Figures 1a, b and c). Moreover, the GPELE significantly suppressed the formation of the ulcers and it was interesting to note the flattening of gastric mucosal folds in rats pretreated with GPELE. It was also observed that protection of gastric mucosa was more prominent in rats pre-treated with 400 mg/kg rhizome extract (Table 1). Furthermore, ethanol-
induced mucosal damage was significantly and dose dependently reduced in the size and severity by pretreatment of the animals with GPELE. The significant inhibition of gastric ulcer in pretreatment with GPELE was comparable with omeprazole which is a standard drug used for curing gastric ulcer.

Histological evaluation of gastric lesions

Histological observation of ethanol induced gastric lesions in ulcer control group pre-treated with CMC only, showed comparatively extensive damage to the gastric mucosa, and oedema and leucocytes infiltration of the submucosal layer (Figure 1d). Rats that received pre-
treatment with GPELE had comparatively better protection of the gastric mucosa as seen by reduction in ulcer area, reduced or absent submucosal edema and leucocytes infiltration (Figure 1e and f). The GPELE has been shown to exert the cytoprotective effects in a dose-dependent manner.

DISCUSSION

Peptic ulcers are caused when the natural balances
between the aggressive factors of acid and pepsin and
defensive mechanisms of mucus, bicarbonate, mucosal
turnover and blood supply (mucosal barrier) are disturbed
(Piper and Stiel, 1986). Baron et al., (1980) have sug-
gested that acid and pepsin are relatively less important
as causative agents and that a defect in the defensive
mechanism of gastric mucosa is the first step toward
ulcer formation. Although in most cases the etiology of
ulcer is unknown, it is generally accepted that it is the
result of an imbalance between aggressive factors and
maintenance of the mucosal integrity through the
endogenous defense mechanism (Piper and Stiel, 1986).

It is known that gastric lesions produced by ethanol
administration appear as multiple-hemorrhagic red bands
of different sizes along the glandular stomach. Ethanol is
commonly used for inducing ulcer in experimental rats; it
leads to intense gastric mucosal damage. Studies
suggest that the ethanol-induced damage to the gastro-
intestinal mucosa starts with microvascular injury, namely
disruption of the vascular endothelium resulting in
increased vascular permeability, edema formation and
epithelial lifting (Szabo et al., 1995). Ethanol produces
necrotic lesions in the gastric mucosa by its direct toxic
effect, reducing the secretion of bicarbonates and pro-
duction of mucus (Marhuenda et al., 1993). Exposure to
ethanol increases the extension of cellular damage in a
dose-dependent way (Mutoh et al., 1990).

Omeprazole is a proton pump inhibitor which has been
widely used as an acid inhibitor agent for the treatment of
disorders related to gastric acid secretion for about 15
years (Li et al., 2004). Omeprazole has substituted
benzimidazoles; it inhibits acid secretion by acting on the
hydrogen-potassium exchanger (H+, K+-ATPase) for the
apical plasma membrane of the gastric mucosa (Satoh
et al., 1989). Omeprazole is highly selective for the proton
pump and undergoes catalyzed conversion into active
form within the acid forming space. The active inhibitors
react with SH (thiol) group of the proton pump, resulting in
inhibition of acid formation (Nagaya et al., 1991).

Oxidative stress plays an important role in the patho-
genesis of various diseases including gastric ulcer, with
antioxidants being reported to play a significant role in the
protection of gastric mucosa against various necrotic
agents (Trivedi and Rawal, 2001). Administration of
antioxidants inhibits ethanol-induced gastric injury in rat
(Ligumsky et al., 1995). GPELE possesses a broad
spectrum of biological activities, and the plant extract has
been shown to contain pharmaceutically active chemical
constituents, such as flavonoids, saponins, tannins, and
terprenoids (Akowuah et al., 2002) and it is speculated
that the gastroprotective effect exerted by GPELE could
be attributed to its antioxidant property. Antioxidant
property of the GBELE may possibly counteract oxidative
damage caused by absolute ethanol toxicity. The
observed anti-ulcerogenic activity may be due to its
antioxidant effects and appears to strengthen the
mucosal barrier, which is the first line of defense against
endogenous and exogenous ulcerogenic agents. Pre-
vious studies have shown that flavonoids may be related to
the antiulcer activity (Hiruma-Lima et al., 2006), and
play a major role in the mechanism of gastroprotection
(La Casa et al 2000). GPELE has been reported to contain
flavonoids (Akowuah et al., 2002) and it could be
conceivable that the anti-ulcer activity of this plant could
be linked to the flavonoids since flavonoids are reported
to protect the mucosa by preventing the forma-tion of
lesions by various necrotic agents (Saurez et al., 1996).
It is well known that many flavonoids display anti-secretory
and cytoprotective properties in different experimental
models of gastric ulcer (Zayachkivska et al., 2005).
Flavonoids possess anti-oxidant properties in addition to
strengthening the mucosal defense system through
stimulation of gastric mucus secretion (Martin et al.,
1994) and flavonoids can scavenge for the reactive
oxygen species (superoxide anions) and free radicals
produced by ethanol. These reactive intermediates are
potentially implicated in ulcerogenicity (Lewis and
Hanson, 1991).

The result of the present study also revealed protection
of gastric mucosa and inhibition of leukocytes infiltration
of gastric wall in rats pretreated with GPELE. Similarly,
Kobayashi et al. (2001) reported that teprenone exerts a
protective effect against mucosal lesions through inhibi-
tion of neutrophil infiltration in the ulcerated gastric tissue
and Shimizu et al. (2000) demonstrated that the reduction
of neutrophil infiltration into ulcerated gastric tissue
promotes the healing of gastric ulcers in rats. Cheng and
Koo (2000) showed that oral administration of plant ex-
tract before ethanol administration significantly decreased
neutrophil infiltration of gastric mucosa and Fujita et al.
(1998) observed that an increase in neutrophil infiltration
into ulcerated gastric tissue delayed the healing of gastric
ulcers in rats. Absolute alcohol would extensively da-
mage the gastric mucosa leading to increased neutrophil
infiltration into the gastric mucosa. Oxygen-free radicals
derived from infiltrated neutrophils in ulcerated gastric
tissues have inhibitory effect on gastric ulcers healing in
rats (Suzuki et al., 1998). Neutrophils mediate lipid
peroxidation through the production of superoxide anions
(Zimmerman et al., 1997). Neutrophils are a major source
of inflammatory mediators and can release potent
reactive oxygen species such as superoxide, hydrogen
peroxide and myeloperoxidase derived oxidants. These
reactive oxygen species are highly cytotoxic and can
induce tissue damage (Cheng and Koo, 2000). Further-
more, neutrophil accumulation in gastric mucosa has
been shown to induce microcirculatory abnormalities
(Bou-Abboud et al., 1988). Suppression of neutrophil
infiltration during inflammation was found to enhance
gastric ulcer healing (Tsukimi et al., 1996). GPELE have
been shown to contain anti-inflammatory activity
((Iskander et al., 2002) and it is speculated that the
gastroprotective effect exerted by this plant extract could
be attributed to its anti-inflammatory activity. This
anti-inflammatory activity could also be a key factor in the prevention of gastric ulcer as reported by Swanker et al., (2005).

In the present study, we observed flattening of the mucosal folds which suggests that gastroprotective effect of GPELE might be due to a decrease in gastric motility. It is reported that the changes in the gastric motility may play a role in the development and prevention of experimental gastric lesions (Garrick et al., 1986; Takeuchi et al., 1987). Relaxation of circular muscles may protect the gastric mucosa through flattening of the folds. This will increase the mucosal area exposed to necrotizing agents and reduce the volume of the gastric irritants on rugal crest (Takeuchi and Nobuhaara, 1985). Ethanol produces a marked contraction of the circular muscles of rat fundic strip. Such a contraction can lead to mucosal compression at the site of the greatest mechanical stress, at the crests of mucosal folds leading to necrosis and ulceration (Mersereau and Hinchey, 1982). The acute toxicity profile of GPELE could be considered favorable, judging from the absence of adverse clinical manifestations in experimental animals after 14 days of observation. It is concluded that the extract has no acute toxicity and that the oral lethal dose for male and female rats is in excess of 5 g/kg.

In conclusion, GPELE could significantly protect the gastric mucosa against ethanol-induced injury. Such protection was shown to be dose dependent as ascertained by the reduction of ulcer areas in the gastric wall as well as the reduction or inhibition of edema and leucocytes infiltration of submucosal layers, and protection was most prominent at a dose of 400 mg/kg rhizome extract. The data obtained confirm the traditional indications for this herb and present a new therapeutic option for the treatment of gastric ailments. The exact mechanism(s) underlying this anti-ulcerogenic effect remain unknown, but it seems that this extract contains pharmacologically active substances with potent antioxidant and anti-inflammatory activity.

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