Protective Immune Response in BALB/c Mice Induced by DNA Vaccine of the ROP8 gene of Toxoplasma gondii

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Abstract. Toxoplasmosis in humans and other animals is caused by the protozoan parasite Toxoplasma gondii. During the process of host cell invasion and parasitophorous vacuole formation by the tachyzoites, the parasite secretes Rhoptyr protein 8 (ROP8), an apical secretory organelle. Thus, ROP8 is an important protein for the pathogenesis of T. gondii. The ROP8 DNA was constructed into a pVAX-1 vaccine vector and used for immunizing BALB/c mice. Immunized mice developed immune response characterized by significant antibody responses, antigen-specific proliferation of spleen cells, and production of high levels of IFN-γ (816 ± 26.3 pg/mL). Challenge experiments showed significant levels of increase in the survival period (29 days compared with 9 days in control) in ROP8 DNA vaccinated mice after a lethal challenge with T. gondii. Results presented in this study suggest that ROP8 DNA is a promising and potent vaccine candidate against toxoplasmosis.

INTRODUCTION

Toxoplasma gondii is an obligate intracellular parasite that is known to invade and replicate in almost nucleated cells including humans; in humans, primary infection can lead to mild fever, lymphadenopathy, lethargy, and sore throat. More severe infection is observed in immunocompromised patients leading to myocarditis, hepatitis, splenomegaly, polymyositis, encephalitis, dermatomyositis, chorioretinitis, and cognitive impairment. In expectant women, the transplacental transmission of the parasite leads to abortion, malformations during the birth of the infant, neurological disorder, loss of sight, and sometimes fetal mortality; in animals, the infection results in abortion generating huge economic losses all over the world and forms the major source of transmission to humans.

Treatment of toxoplasmosis is difficult caused by toxic effects of the drugs and possible reinfection. Vaccination is considered the most potent and effective means of fighting the infection. By far the only existing vaccine for toxoplasmosis is the live, attenuated formulation using T. gondii S48 strain. However, this attenuated vaccine is considered inappropriate for use in livestock because of the risk of reversion of the infection by the most proliferative form of the parasite.

The DNA vaccination method that uses the plasmid vector to express the target antigen. They are known to induce cellular and humoral immune response in animal models and considered to be safe, efficient, and cost effective. Immunization studies with plasmid DNA have been known to induce both cellular and humoral responses in T. gondii.

The rhoptry proteins are secreted during host cell invasion of the parasite involved in the parasitophorous vacuole formation and associated with the host cell mitochondria. The Rhoptyr protein 8 (ROP8) is one of the most abundant proteins belonging to the ROP2 family of proteins and contains a conserved Serine/Threonine kinase domain. However, rhoptry proteins are degenerate in key positions normally invariant in active eukaryotic kinases and catalytically inactive. Several rhoptry genes such as ROP1, 2, 16, and 18 have been evaluated for their potential as vaccine candidates against toxoplasmosis. However, the vaccine potential of ROP8 has not been evaluated. Therefore, in this study we evaluated the vaccine potential of ROP8 DNA by evaluating the vaccine-induced protective immune response against toxoplasmosis in mouse model.

MATERIALS AND METHODS

Mice and parasites. Six- to 8-week-old female BALB/c mice obtained from the University of Malaya, Animal Experimental Center were used in this study. Use of animals was approved by the Animal Care and Use Committee of the University of Malaya, Faculty of Medicine. Highly virulent tachyzoites (RH-strain) of T. gondii were maintained by successive intraperitoneal passage in mice. Parasites were collected from the intraperitoneal fluids, washed with phosphate buffered saline (PBS), and filtered through 3 μm polycarbonate filters. The filtered suspension was then centrifuged at 1,500 rpm for 10 min and resuspended in sterile PBS for further use. The freshly isolated parasites were also subjected to repeated freeze-thaw cycles to extract total lysate antigen (TLA).

Cloning of ROP8 in eukaryotic expression vector pVAX-1. The genomic DNA of tachyzoites (RH-strain) was isolated using a kit from Qiagen (Valencia, CA). The 1,728 bp ROP8 gene (GenBank ID: AF011377) was amplified using gene-specific primers (forward primer 5'CCCAAGCTTATGATCACCCATACGGATATTTTCTGTGTTACG 3' and reverse primer 5'CAGGATTCTCATGCCGTTCCATCAG 3' containing the HindIII and EcoR1 restriction sites as underlined). The polymerase chain reaction (PCR) condition used was 95°C for 10 min for initial denaturation before 30 cycles of amplification, which consisted of denaturation at 94°C for 1 min, annealing at 60°C for 1 min, and elongation at 72°C for 2 min. Final extension of the reaction was carried out at 72°C for 10 min. The ROP8 gene was digested with HindIII and EcoR1 and ligated into the pVAX-1 vector (Invitrogen, USA) digested with the same restriction enzymes. The resulting construct ROP8-pVAX-1 was used for the vaccination studies. The ROP8 gene was also cloned into the
pVAX-1-GFP vector and the resulting construct ROP8-pVAX-1-GFP was used for in vitro expression of ROP8 in Chinese hamster ovary (CHO) cells (Figure 1). The pVAX-1 vector meets all the guidelines required for the expression of target genes in animal models and fulfills the requirements by the U.S. Food and Drug Administration (FDA) for formulation of DNA vaccines; the green fluorescent protein (GFP) gene was incorporated into the pVAX-1 vector to generate pVAX-1-GFP and ROP8 gene is fused to the N-terminus of GFP. Positive recombinant plasmids (ROP8-pVAX-1 and ROP8-pVAX-1-GFP) were extracted using a commercial kit (EndoFree Plasmid free Giga kit, Qiagen). The DNA concentration was quantified using NanoDrop at 260 and 280 nm and sent to a commercial laboratory (GenScript Inc., Piscataway, NJ) for sequencing.

**In vitro expression of ROP8-pVAX-1-GFP in CHO cells.** The CHO cells were transfected with ROP8-pVAX-1-GFP plasmid using TurboFect (Fermentas Inc., Glen Burnie, MA), lipid-based transfection reagent. The transfected cells were observed under an inverted florescence microscope (Olympus, Hicksville, NY) at 24, 48, and 72 h post transfection for the expression of the GFP-tagged recombinant protein. The CHO cells transfected with pVAX-1-GFP vector was used as the control. The cells were harvested and total proteins were extracted and separated on a sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) gel. The separated proteins were also transferred by electrophotography onto polyvinylidene difluoride (PVDF) membranes (Bio-Rad, USA) under a reducing condition. Blocking of the membranes was done by incubation in Tris-buffered saline (TBS), containing 5% skimmed milk overnight at 4°C. The membranes were washed three times with TBS-t (TBS containing 0.2% Tween-20), and then probed with GFP monoclonal antibody (Invitrogen, diluted 1:1000) or anti-T. gondii positive human sera diluted (1:250) in TBS containing 2.5% skimmed milk for 2 hours with constant shaking at room temperature. After three additional washes, the membrane was treated for 1 hour at room temperature with 1:2500 diluted biotin-labeled anti-mouse immunoglobulin G (IgG) and anti-human IgG (KPL Inc., Gaithersburg, MA), followed by streptavidin-AP (Invitrogen Corp., USA). Finally, the protein band was revealed using NBT/BCIP (Sigma Chemical CO., USA) as the chromogenic substrate. The color was allowed to develop at room temperature in the dark. The membrane was dried and photographed.

**DNA immunization and challenge.** Three groups of mice (15 mice per group) were immunized with 100 μg of plasmid DNA suspended in 100 μL sterile PBS by bilateral intramuscular injection into the quadriceps and boosted twice with the same dose at 2-week intervals. Group I was injected with ROP8-pVAX-1, group II with empty pVAX-1 vector as control, and group III with PBS as control. One day before the challenge, blood was collected from the tail vein of each animal and sera were stored at -20°C; twelve mice from each group were challenged intraperitoneally with 1 x 10³ tachyzoites. These experiments were repeated twice. The mice sera were tested for presence of ROP8-specific antibodies by western blotting. The TLA proteins separated by SDS-PAGE were transferred by electroblotting as previously described. The membranes were probed with mice sera (diluted 1:250) and later treated with 1:2500 diluted biotin-labeled anti-mouse IgG followed by streptavidin-AP. The protein band was detected using NBT/BCIP substrate.

**Cell proliferation assay.** Spleens of three mice from each group were aseptically removed 2 weeks after the last booster injection and single cell suspensions were prepared individually. After lysing the red blood cells with ACK lysis solution, splenocytes were resuspended in RPMI medium supplemented with 10% fetal calf serum, 0.5% gentamicin, and 1% glutamine. Cells were plated in 96-well Costar plates at a density of 2.5 x 10⁶ cells per well and cultured with TLA of tachyzoites (10 μg/mL) and concanavalin A as positive control (5 μg/mL; Sigma) and medium alone as negative control at 37°C with 5% CO₂ for 72 h. The proliferative activity was measured using BrdU colorimetric assay (Roche, Indianapolis, IN), according to the manufacturer’s instructions. The stimulation index was calculated as the ratio of the average OD₅₇₀ value of wells containing antigen-stimulated cells to the average OD₅₇₀ value of wells containing only cells with medium. These experiments were repeated twice.

**Cytokine assays.** Culture supernatants from splenocytes stimulated with the tachyzoite lysates were harvested 72 h after stimulation and assayed for levels of interleukin-4 (IL-4) and gamma-interferon (IFN-γ) using a commercial ELISA Kit (BD Science, San Jose, CA). Cytokine concentrations were determined by reference to standard curves constructed with known amounts of mouse recombinant IFN-γ, and IL-4.

**Statistical analysis.** Data were analyzed using SPSS software (SPSS, Inc., Chicago, IL) and their statistical significance

![Figure 1](Image)
was determined by the analysis of variance test. A value of \( P < 0.05 \) was considered as significant.

**RESULTS**

**Expression of ROP8-pVAX-1-GFP plasmid in CHO cells.**
Expression of ROP8-pVAX-1-GFP plasmid was evaluated in CHO cells by fluorescence microscopy at 72 h after transfection (Figure 2). The results showed that CHO cells can be transfected with ROP8-pVAX-1-GFP and pVAX-1-GFP plasmids as evidenced by the green fluorescence. Subsequently, we confirmed the identity of the recombinant ROP8 (rROP8) by western blotting using GFP mAb or the serum from a toxoplasmosis-positive patient. The GFP mAb and patient serum detected the GFP tagged rROP8 at 92 kDa (Figure 3A and B).

**Humoral immune response of ROP8-pVAX-1 in mice.** The sera obtained from mice (\( N = 15 \)) of all the groups one day before the challenge were analyzed individually for the presence of specific ROP8 antibody by western blotting using TLA of the tachyzoites. We observed that only the serum from mice vaccinated with ROP8-pVAX-1 reacted with a protein of 65 kDa in the TLA of tachyzoites. The estimated size of the native ROP8 is 65 kDa (Figure 4). Only two strips for each group are presented.

**Lymphoproliferative immune response.** Spleen cells from mice immunized with ROP8-pVAX-1 2 weeks after the final injection showed significant proliferative response to TLA (1.7 ± 0.06) compared with the pVAX-1 control or PBS (\( P < 0.05 \)) (Figure 5).

**Cytokine production.** The TLA stimulated splenocyte culture supernatants from mice immunized with ROP8-pVAX-1 or pVAX-1 were evaluated for the production of IL-4 and IFN-\( \gamma \). The result showed that splenocytes from mice immunized with ROP8-pVAX-1 secreted significantly high levels of IFN-\( \gamma \) (816 ± 26.3 pg/mL) compared with spleen cells cultured from mice immunized with pVAX-1 (48 ± 10.8) or PBS (45 ± 6.6). Only a low level of IL-4 was detected in the culture supernatant of splenocytes from mice immunized with ROP8-pVAX-1 (148 ± 18.3) but was significantly higher compared with mice immunized with pVAX-1 (50 ± 13.6) or PBS (47 ± 6.1) (Figure 6).

**Analysis of protective immunity against T. gondii challenge.** To evaluate the level of protection induced by ROP8-pVAX-1...
immunization in mice against *T. gondii* infection, mice were challenged with live tachyzoites (RH strain). The survival curve for the three groups of mice (Figure 7) revealed that there was no difference in survival between the mice immunized with pVAX-1 and PBS. Average survival time was 9 days from the time of challenge. Mice immunized with ROP8-pVAX-1 DNA had a 100% survival rate until Day 9, whereas all the control mice died. On top of that, mice immunized with ROP8-pVAX-1 DNA showed a significant increase in the survival time (29 days, *P* < 0.05).

**DISCUSSION**

The data presented in this study show that ROP8 DNA vaccination developed significant humoral and cell-mediated immune response and confers significant protection in mice against a lethal challenge of the parasite.

Before performing the DNA vaccine study, we confirmed that the ROP8 gene can be expressed in eukaryotic cells by transfecting CHO cells. The recombinant rROP8-GFP expressed in CHO cells had an apparent molecular mass of 92 kDa that was 27 kDa greater than that of the native ROP8 protein found in the tachyzoite lysate. The difference in the molecular weight is caused by the fusion of the GFP protein (24 kDa) to the rROP8 along with the post-translational modifications associated with the recombinant protein. The identity of rROP8 in CHO cells was further confirmed using GFP mAb, again showing a single band at the same size. Mice immunized with the ROP8 DNA did not show any signs of adverse reaction. These immunized mice developed significant levels of anti-ROP8 IgG antibodies that recognize native ROP8 protein found in TLA of *T. gondii* in western blots. Our results thus show that DNA vaccination with ROP8 is immunogenic and generates specific antibodies.

We then asked the question, if this DNA immunization can confer protection against a challenge infection. Various *T. gondii* proteins such as the micronemal protein, dense granular protein, rhoptry proteins, matrix protein, and surface antigens have been evaluated as potential vaccine candidates for toxoplasmosis. Results presented in this study show that ROP8 is also a potential vaccine candidate. Mice immunized with ROP8-pVAX-1 survived for a prolonged period of time (29 days) after a challenging infection compared with the other groups.
control groups. Previous studies using ROP16 and ROP18 DNA vaccination trials also gave a similar survival time of 21.6 ± 9.9 and 27.9 ± 15.1 days, respectively, in mice. Our studies show that compared with ROP16 and ROP18, the ROP8 DNA vaccination confers a higher degree of protection.

The IFN-γ dependent Th1-predominant immune response appears to be critical for protection against T. gondii. The ROP8 vaccinated animals in our studies showed that antigen-specific spleen cells secreted high levels of IFN-γ. We also found that there is a significant correlation between the presence of high levels of IFN-γ secreting cells and degree of protection conferred. Thus, our findings are in agreement with previous reports that Th1-mediated responses are highly protective in T. gondii infection.

This study showed that a DNA vaccine encoding the T. gondii ROP8 protein is a simple and potent vaccine that elicits specific immune responses, and provides significant protection against a virulent strain of T. gondii. Therefore, ROP8 can be considered as a potential vaccine candidate against toxoplasmosis. It may be possible to provide complete protection against T. gondii infection by combining ROP8 with other immunogenic proteins as previously described, such as ROP16, ROP18, MIC3, SAG1, and GRA4.

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