Antifungal and antibacterial properties of three medicinal plants from Malaysia

Gwee Pei Shing¹, Chen Li Wen¹, Tan Syu Wei¹, Ong Hean Chooi², Khoo Kong Soo³ and Sit Nam Weng¹*

¹Department of Biomedical Science, Faculty of Science, Universiti Tunku Abdul Rahman, Bandar Barat, 31900 Kampar, Perak, Malaysia
²Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia
³Department of Chemical Science, Faculty of Science, Universiti Tunku Abdul Rahman, Bandar Barat, 31900 Kampar, Perak, Malaysia

ABSTRACT: Introduction: The increase of opportunistic fungal infections and the escalation of bacterial resistance have seriously reduced the efficacy of chemotherapeutic agents available. Thus the search for new antimicrobial agents from natural sources such as medicinal plants becomes necessary. Methods: The aerial parts of Diplazium esculentum and Sechium edule, and the fruits of Solanum muricatum were used, and extracted sequentially using hexane, chloroform, ethyl acetate, ethanol, methanol, and water. The extracts were then evaluated, in triplicate, against a panel of 12 medically-important microorganisms for microbistatic and microbicidal activities using colorimetric broth microdilution methods. Results: The total percentage yield obtained were 1.20%, 1.84% and 3.53% (w/w, based on fresh weight) for D. esculentum, S. edule and S. muricatum, respectively. All plant extracts showed antifungal activity with 66% and 49% of the bioassays demonstrating fungistatic and fungicidal activity, respectively. Two yeasts, Cryptococcus neoformans and Issatchenkia orientalis were found to be susceptible to all extracts. The lowest minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) was exhibited by the hexane extracts of S. edule and S. muricatum against C. neoformans, both with values of 0.08 mg/mL. In the antibacterial screening assays, 49% of the bioassays exhibited bacteriostatic activity while only 21% of them showed bactericidal activity. The lowest MIC and minimum bactericidal concentration (MBC) was recorded for the hexane extract of S. muricatum against Bacillus cereus and Klebsiella pneumoniae, both with values of 0.31 mg/mL. The susceptibility of bacteria towards the plant extracts evaluated was species-dependent, with the susceptibility indices ranging from 0% for Escherichia coli to 72% for Pseudomonas aeruginosa. Conclusions: The results from this study show that extracts from these plants have significant antimicrobial activity, which corroborates their use in traditional medicine.

KEYWORDS: bacteriostatic, bactericidal, broth microdilution, extraction, fungistatic, fungicidal, Diplazium esculentum, Sechium edule, S. muricatum

INTRODUCTION

For centuries, herbal remedies derived from medicinal plants have been a major source of medicine for the treatment and prevention of ailments.[2,3] For example, the use of pumpkin (Cucurbita moschata) seeds and cranberry (Vaccinium macrocarpon) juice to treat urinary tract infections, while species such as garlic (Allium sativum) and tea tree (Melaleuca alternifolia) are used as broad-spectrum antimicrobial agents.[2,3]

The increase of opportunistic fungal infections and the escalation of bacterial resistance, particularly multi-drug resistance have seriously reduced the efficacy of many chemotherapeutic agents. The clinical usefulness of some antibiotics may be diminished within a short time due to the over-prescription and misuse of antibiotics.[4] Fungal infections remain the fourth-leading cause of life-threatening infections in hospitals, in part as the result of alterations in immune status associated with Acquired Immune Deficiency Syndrome (AIDS) epidemic, cancer chemotherapy and organ or bone marrow transplantation.[5] The most common fungal infection agents are those ubiquitous colonizers such as Candida spp., Cryptococcus and Aspergillus spp. with an overall mortality for invasive diseases of 25–50%.[6–9]

In Malaysia, the ‘vegetable fern’ (D. esculentum) and ‘chayote’ (S. edule), are usually eaten cooked in various dishes,
while the fruits of “pepino” (S. muricatum) are mostly consumed as a dessert.[8,10] D. esculentum (vegetable fern) is used traditionally to treat expectoration of blood, fever, dermatitis, measles, coughs and taken as a tonic by woman after childbirth.[8,12] S. edule (chayote) is used as a folk medicine in the treatment of arteriosclerosis, calcifications in the urinary system, hypertension and fever. The flesh of the chayote fruit is applied as a poultice on inflammations and wounds, while the decoction and juice are taken for their diuretic effect, and to treat hypertension and pulmonary ailments.[8,12,13] S. mural catum (sweet pepino) is used as a diuretic, and for the treatment of hypotension.[14,15]

This study was conducted to evaluate the antimicrobial activities of vegetable fern, chayote and sweet pepino against a panel of pathogenic bacteria (Staphylococcus aureus, Bacillus cereus, Klebsiella pneumoniae, Pseudomonas aeruginosa, Escherichia coli, and Acinetobacter baumannii) and fungi (Candida albicans, Candida parapsilosis, Issatchenka orientalis, Cryptococcus neoformans, Aspergillus brasiliensis and Trichophyton mentagrophytes).

MATERIALS AND METHODS

Chemicals and reagents

The following chemicals and reagents were used: Hexane (Mallinckrodt Chemicals, USA), chloroform (System, USA), ethyl acetate (R&M, UK), ethanol (PROCHEM, USA), methanol (RCI Labscan, Thailand), amphotericin B and p-iodonitrotetrazolium violet (Sigma-Aldrich, USA), chloramphenicol (Amresco, USA), potato dextrose agar (PDA) and sodium hydroxide pellets (Merck, Malaysia), Mueller-Hinton agar (MHA) and Mueller- Hinton broth (MHB) (Scharlau Microbiology, Spain), RPMI-1640 medium supplemented with glutamine and phenol red, without bicarbonate (MP Biomedicals, France) and 3-(N-morpholino)propanesulfonic acid (MOPS) (Calbiochem, Germany).

Strains tested

Bacterial strains. Staphylococcus aureus (ATCC 6538), Bacillus cereus (ATCC 11778), Klebsiella pneumoniae (ATCC 13883), Pseudomonas aeruginosa (ATCC 27853), Escherichia coli (ATCC 35218) and Acinetobacter baumannii (ATCC 19606) were purchased commercially from ATCC. The cultures were maintained on MHA at 4°C.

Fungal strains. Candida albicans (ATCC 90028), Candida parapsilosis (ATCC 22019), Issatchenka orientalis (ATCC 6258), Cryptococcus neoformans (ATCC 90112), Aspergillus brasiliensis (ATCC 16404) and Trichophyton mentagrophytes (ATCC 9533) were purchased commercially from ATCC. The microorganisms were maintained on PDA at 4°C.

Plant materials

The aerial part of S. edule and the fruits of S. muricatum were purchased from a marketplace in Cameron Highlands, Malaysia while the aerial part of D. esculentum was obtained from a wet market in Kampar, Malaysia. The identification of these plants was ascertained by a co-author of this paper, who is a botanist (H.C. Ong). Voucher specimens of D. esculentum (UTAR/FSC/10/023) and S. edule (UTAR/FSC/10/022) were prepared and deposited at the Faculty of Science, Universiti Tunku Abdul Rahman. No voucher specimen was prepared for the fruit of S. muricatum.

Preparation of extracts

Fresh plant materials were washed thoroughly using tap water. The collected parts of fresh plant samples were blended and immersed in the appropriate solvent. Samples of D. esculentum (1214 g), S. edule (1200 g) and S. muricatum (1000 g) were sequentially extracted with hexane, chloroform, ethyl acetate, ethanol, methanol and distilled water at room temperature with agitation (120 rpm) using an orbital shaker (IKA- Werke KS 501, Germany). Two cycles of extractions were performed for each solvent. The solvent was filtered, evaporated in a rotary evaporator (BUCHI Rota-vapor R205, Switzerland) at 40°C. The water extracts were lyophilized using a freeze-dryer (Martin Christ Alpha, UK). Yields of extracts are presented in Figure 1. For bioassay, the extracts were re-dissolved in methanol: water solution (2:1, v/v) at a concentration of 10 mg/mL, filtered using 0.45 μm nylon syringe filters and stored at –20°C prior analyses.

Antimicrobial screening

A colorimetric broth microdilution method using 96-well round bottom microplates was employed for antimicrobial activities screening of the extracts with modifications.[16] The test was conducted in serially by two-fold descending

![Figure 1](image-url). The percentage of yield of various extracts from three medicinal plants, D. esculentum, S. edule and S. muricatum.
concentrations of extracts and antibiotics, with the concentration ranging from 2.50 to 0.02 mg/mL for the plant extracts, 128 to 1 μg/mL for chloramphenicol and 8 to 0.06 μg/mL for amphotericin B. Growth, sterility (medium only) and negative (extracts only) controls were included. The colorimetric indicator, p-iodonitrotetrazolium violet (INT) was prepared in distilled water at the concentration of 0.4 mg/mL. To indicate the antimicrobial activity, INT was added after incubation. A colour change (from colourless to red) was indicative of a positive result. The concentration of extract at which the colour remains clear was recorded as the minimum inhibitory concentration (MIC) value. The minimum bactericidal concentration (MBC) or minimum fungicidal concentration (MFC) was obtained by inoculating 20 μL of the preparation that showed no evidence of bacterial or fungal growth during the MIC determination assays on MHA or PDA, respectively. The lowest concentration of extract which inhibited was recorded as the MBC or MFC value. The test was performed in triplicate.

**Screening for antibacterial activity**

The standard method was used to prepare broth medium and bacterial culture. Three to five healthy colonies of bacteria grown on MHA (24 h old, 37°C) were transferred to MHB. The bacterial concentration was adjusted to the optimal absorbance value (OD = 0.08 to 0.10) at 625 nm, and subsequently diluted with MHB to 1 × 10^6 CFU/mL. The final inoculum (100 μL) of bacteria in each well of the microplate was 5 × 10^5 CFU/mL. The microplates were covered and incubated for 24 h at 37°C.

**Screening for antifungal activity**

Preparation of broth medium and inoculum suspensions were based on the CLSI/NCCLS guidelines. Inoculum suspensions were prepared from fresh, mature cultures (48 h for Candida spp.; 72 h for C. neoformans; 7-day-old for A. brasiliensis and T. mentagrophytes) grown on PDA. The suspensions were mixed for 15 s and adjusted to the optimal absorbance value (OD = 0.12–0.15 for Candida spp. and C. neoformans; OD = 0.09–0.11 for A. brasiliensis and OD = 0.15–0.18 for T. mentagrophytes) at 530 nm. Further dilution in sterile distilled water was performed in order to obtain the required final working inoculum (1–5 × 10^3 CFU/ml for Candida spp.; 1–5 × 10^4 CFU/ml for C. neoformans; 0.4–5 × 10^4 CFU/ml for A. brasiliensis and 1.2–6 × 10^4 CFU/ml for T. mentagrophytes). After addition of the plant extracts and antibiotics into the 96-well microplates using the same dilution technique, the microplates were incubated at 35°C for 48 h for Candida spp.; 72 h for C. neoformans and A. brasiliensis; and at 28°C for 7 days for T. mentagrophytes.

**RESULTS**

The percentage yields (w/w, based on fresh weight) obtained from the sequential extraction of D. esculentum, S. edule and S. muricatum are presented in Figure 1. Solanum muricatum showed the highest total percentages of yields (3.53%) followed by S. edule (1.84%), while the lowest yield was obtained from D. esculentum (1.20%).

Eighteen extracts from three medicinal plants were tested for antimicrobial activity against two Gram-positive bacteria, four Gram-negative bacteria, four yeasts and two molds using colorimetric broth microdilution methods. The results of the antibacterial and antifungal activities are shown in Tables 1 and 2, respectively. In the antibacterial screening assays, 49% of the assays exhibited bacteriostatic activity while only 21% of them showed bactericidal activity. The lowest MIC and MBC were recorded for the hexane extract of S. muricatum against Bacillus cereus and K. pneumonia, both with values of 0.31 mg/mL. The water extract of D. esculentum, the methanol extract of S. edule, and the ethanol and methanol extracts of S. muricatum showed selective inhibitory activity against P. aeruginosa, with the MIC range of 0.31 to 1.25 mg/mL. None of the extracts showed inhibitory activity against E. coli. All the plants showed antifungal property with 66% and 49% of the bioassays demonstrated fungistatic and fungicidal activities, respectively. The lowest MIC and MFC values (both 0.08 mg/mL) obtained from the hexane extracts of S. edule and S. muricatum against C. neoformans. The ethanol, methanol and water extracts of S. edule and S. muricatum, and the water extract of D. esculentum exhibited selective inhibitory activity against C. neoformans and I. orientalis, with MIC values ranging from 0.16–2.50 mg/mL.

The bacterial susceptibility index (BSI) and the fungal susceptibility index (FSI) are shown in Figures 2 and 3, respectively. The susceptibility of bacteria towards the plant extracts
evaluated was species-dependent, with the susceptibility indices ranged from 0% for *E. coli* to 72% for *P. aeruginosa*. All the extracts exhibited antifungal activity against at least two strains of fungi. Two yeasts, *C. neoformans* and *I. orientalis* were found to be 100% susceptible to all the plant extracts with MIC values ranging from 0.08 to 2.50 mg/mL. *Aspergillus brasiliensis* was found to be the most insensitive among the tested fungal strains with a susceptibility index of 33%.

### DISCUSSION

In this study, a total of 18 extracts isolated from three plants, *D. esculentum*, *S. edule* and *S. muricatum* were subjected to antimicrobial activity screening. Although these medicinal plants have been traditionally used as herbal remedies, there has been relatively few studies regarding their effects on human pathogens.

The extracts are regarded to have strong inhibitory effects if the MIC values is 0.5 mg/mL and below; moderately inhibitory if the MIC value is between 0.6 and 1.5 mg/mL; and weak inhibitory if the MIC value is 1.6 mg/mL and above.
2.50 1.25 0.08 0.63 0.08–0.16 – – – 0.01

The present study

NA – NA – 0.31 0.63 0.63 0.63 0.0001 0.31–0.63 2.50 0.31–0.63 0.31 NA 0.31

Molds

T. mentagrophytes

The MIC values obtained in this study indicate

NA – NA – 0.31 0.63 0.63

the low

NA NA NA NA 0.31–0.63

0.31–0.63

0.31

0.63

0.31

0.008

NA 0.63

0.001

0.63

0.63–1.25

0.63

0.63

0.004

0.0001

Yeasts

C. albicans

I. orientalis

A. brasiliensis

T. mentagrophytes

C. parapsilosis

I. orientalis

C. parapsilosis

C. neoformans

C. neoformans

I. orientalis

C. neoformans

S. muricatum

S. muricatum

S. edule

S. edule

D. esculentum

D. esculentum

and weak inhibitory if the MIC value is 1.6 mg/mL or more.[29] The MIC values obtained in this study indicate that the tested plant extracts are generally more potent against fungi than bacteria. Based on the antimicrobial assays, 23.1% and 2.8% of the bioassays showed strong inhibition against fungi and bacteria, respectively; with MIC values ranging from 0.08 to 0.31 mg/mL (Tables 1 and 2). The antifungal activity of the extracts was more pronounced on the yeast strains, particularly on C. neoformans and I. orientalis, both with FSI of 100% (Figure 3).

Yeasts (mainly Candida spp.) are the third most common cause of intravascular catheter-related infection, with the second highest colonization-to-infection rate and the overall highest crude mortality.[21] I. orientalis is commonly implicated in urinary tract infections in immuno-compromised patients[22,23] while C. neoformans ranks as one of the most common infectious agents that causes human meningoencephalitis (cryptococcosis).[27] The present study shows that the overall antifungal activity screening results are indicative of the potential of these plant extracts as effective medicaments in the treatment of fungal infectious diseases.

The antimicrobial activities of these plant extracts were found to decrease with increasing polarity of the extracts. According to the antibacterial assays, the lowest MIC and MBC was recorded for the hexane extract of S. muricatum against Bacillus cereus and K. pneumoniae, both with values of 0.31 mg/mL. Based on the antifungal assays, the lowest MIC and MFC was exhibited by the hexane extracts of S. edule and S. muricatum against C. neoformans, both with values of 0.08 mg/mL. The hexane extract of S. muricatum also exhibited strong inhibition against the filamentous fungi, A. brasiliensis and T. mentagrophytes with MIC values of 0.31 and 0.16 mg/mL,
respectively. The results indicate that the non-polar compounds (hexane extracts) had greater antimicrobial activity compared to the more polar compounds extracted by ethanol, methanol and water extracts.

Previous studies demonstrated that ascorbic acid, phenolic acids and flavonoids isolated from the fruits of *S. murrichianum* (pepino) exhibited anti-oxidative, anti-inflammatory and anti-glycative protection in diabetic mice.[26] The antioxidant activity of the ripe pepino fruit was reported to be largely due to polyphenols.[25] The anti-tumor effect of pepino fruits has been reported by Ren & Tang[26] but the active compounds that are responsible for its anti-tumor activity remains to be identified. Eight flavonoids (vicenin-2, apigenin-6-C-β-d-glucopyranosyl-8-C-β-d-apiofuranoside, vitexin, luteolin-7-O-rutinoside, luteolin-7-O-β-d-glucopyranoside, apigenin-7-O-rutinoside, chrysoeriol-7-O-rutinoside, and diosmetin-7-O-rutinoside) have been isolated from the aerial parts of *S. edule.[27] S. edule leave extracts have been reported to possess broad-spectrum antimicrobial activity against *E. coli, K. pneumoniae, Proteus mirabilis, Enterobacter cloacae, Serratia marcescens, Morganella morganii, A. baumannii, P. aeruginosa, Stenotrophomonas maltophilia, Candida spp. and Aspergillus spp.[33]

All the extracts of *D. esculentum*, from non-polar to polar extracts (hexane, chloroform, ethyl acetate, ethanol, methanol and water) showed strong inhibition against *C. neoformans* and *I. orientalis*, with MIC and MFC values ranging from 0.08 to 0.31 mg/mL (Table 2). These results agree well with another study which reported the antifungal activity of the methanolic extract from *D. esculentum.[28] However, the antifungal activity of *D. esculentum* reported was limited to a few species (*A. niger, Rhizopus stolonifer* and *C. albicans*). The results showed the inhibitory property of the extracts was weak, with MIC values ranging from 50 mg/mL to 100 mg/mL.

In the antibacterial screening assays, 66.7% of the extracts from *D. esculentum* exhibited bacteriostatic activity (Table 1). Sakunpak and Panichayupakarananta[29] reported that extracts of *D. esculentum* did not exhibit inhibitory activity against gastrointestinal pathogenic bacteria, including *E. coli* (ATCC 25922) and *S. aureus* (ATCC 25923). The data reported show that extracts of *D. esculentum* have substantial antimicrobial activity against medically-important microorganisms, which corroborates its use in traditional medicine for the treatment of skin infections such as dermatitis and measles.

The antimicrobial activity profile of the three plants indicated that *E. coli* (BSI = 0%) was the least susceptible bacterium (Figure 2) to the plant extracts evaluated. *E. coli* is one of the most frequent causes of bacterial infections, including cholecystitis, bacteremia, cholangitis, urinary tract infection, diarrhoea, neonatal meningitis, and ulcerative colitis.[30–32] The *E. coli* (ATCC 35218) strain used in this study is an ampicillin-resistant strain.[33] *E. coli* is among the Gram-negative bacteria that develop multi-drug resistance.[34]

**CONCLUSION**

All the plants investigated possessed antifungal and antibacterial properties against human pathogens. The results of this study corroborate the usage of these plants in traditional medicine. Further studies will be carried out to isolate and identify the active compounds.

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**REFERENCES**